

地榆中化学成分的研究

范琳资, 黄 静, 王晓童, 吴承翠, 姜 爽, 杨春娟*

哈尔滨医科大学药学院 药物分析与分析化学教研室, 哈尔滨 150081

摘要: 蔷薇科植物地榆 (*Sanguisorba officinalis* L.) 的 70% 乙醇提取物中共提取分离得到 20 个化合物, 经过理化和波谱分析分别鉴定为地榆素 H-4 (1)、3,4-二羟基-5-甲氧基苯甲酯 (2)、没食子酸 (3)、没食子酸甲酯 (4)、儿茶素 (5)、表儿茶素 (6)、没食子儿茶素 (7)、表没食子儿茶素 (8)、表没食子儿茶素没食子酸酯 (9)、木犀草素-7-*O*- β -D-葡萄糖苷 (10)、坡模酮酸 (11)、2 α -羟基齐墩果酸 (12)、白桦脂酸 (13)、齐墩果酸 (14)、地榆皂苷 II (15)、1,4,6-三-*O*-没食子酰基- β -D-吡喃葡萄糖 (16)、methyl-6-*O*-galloyl- β -D-glucopyranoside (17)、(*E*)-7-hydroxy-3,7-dimethyl-2-octenyl-6-*O*- α -L-arabinofuranosyl- β -D-glucopyranoside (18)、7-hydroxy-3,7-dimethyloctyl-6-*O*- α -L-arabinofuranosyl- β -D-glucopyranoside (19)、n-butyl- β -D-glucopyranoside (20)。其中, 化合物 9、20 为首次从地榆属植物中分得。研究结果对地榆中主要化学成分进行了进一步确证。

关键词: 地榆; 三萜类; 黄酮类; 分离; 结构鉴定

中图分类号: Q946.91

文献标识码: A

文章编号: 1001-6880(2020)Suppl-0015-08

DOI: 10.16333/j.1001-6880.2020.S.003

Research on chemical constituents isolated from *Sanguisorba officinalis* L.

FAN Lin-zi, HUANG Jing, WANG Xiao-tong, WU Cheng-cui, JIANG Shuang, YANG Chun-juan*

Department of Pharmaceutical Analysis and Analytical Chemistry, College of Pharmacy, Harbin Medical University, Harbin 150081, China

Abstract: *Sanguisorba officinalis* L. (the dried root of *Sanguisorba*, Rosaceae) is a traditional medicinal herb that is widely distributed in Europe and Asia. It has been officially recorded in the Chinese Pharmacopoeia (2015), possesses activities such as detoxification, analgesia and hemostasis, and has been used for the prevention and treatment of various diseases in tradition, including burns and scalds, bleeding hemorrhoids, metrostaxis, bleeding wounds, hematochezia, and swollen carbuncles. In order to better evaluate the quality of *S. officinalis*, the chemical constituents of the 70% EtOH extract was investigated. After physicochemical and spectral analysis, 20 compounds were isolated and identified: sanguin H-4 (1), 3,4-dihydroxy-5-methoxybenzyl ester (2), gallic acid (3), methyl gallate (4), catechin (5), epicatechin (6), gallic acid (7), epigallocatechin (8), epigallocatechin gallate (9), luteolin-7-*O*- β -D-glucoside (10), pomonic acid (11), 2 α -hydroxyl oleanolic acid (12), betulinic acid (13), oleanolic acid (14), ziyuglycoside II (15), 1,4,6-tri-*O*-galloyl- β -D-glucopyranose (16), methyl-6-*O*-galloyl- β -D-glucopyranoside (17), (*E*)-7-hydroxy-3,7-dimethyl-2-octenyl-6-*O*- α -L-arabinofuranosyl- β -D-glucopyranoside (18), 7-hydroxy-3,7-dimethyloctyl-6-*O*- α -L-arabinofuranosyl- β -D-glucopyranoside (19), n-butyl- β -D-glucopyranoside (20). Among them, compounds 9 and 20 were isolated from the genus *Sanguisorba* for the first time. The results provided a material basis for the quality evaluation of *S. officinalis*.

Key words: *Sanguisorba officinalis* L.; triterpenoids; flavonoids; separation; structure identification

地榆 (*Sanguisorba officinalis* L.) 为蔷薇科地榆属植物地榆 *S. officinalis* 或长叶地榆 *Sanguisorba of-*

ficialis L. var. *longifolia* (Bert.) Yü et Li 的干燥根, 前者盛产于我国南北各地, 后者主要产于安徽、浙江、江苏、江西等地, 又习称“绵地榆”, 为《中国药典》(2015 版) 收载药物。其味苦、酸、涩, 性微寒, 归肝、大肠经。具有凉血止血, 清热解毒, 消肿敛疮的

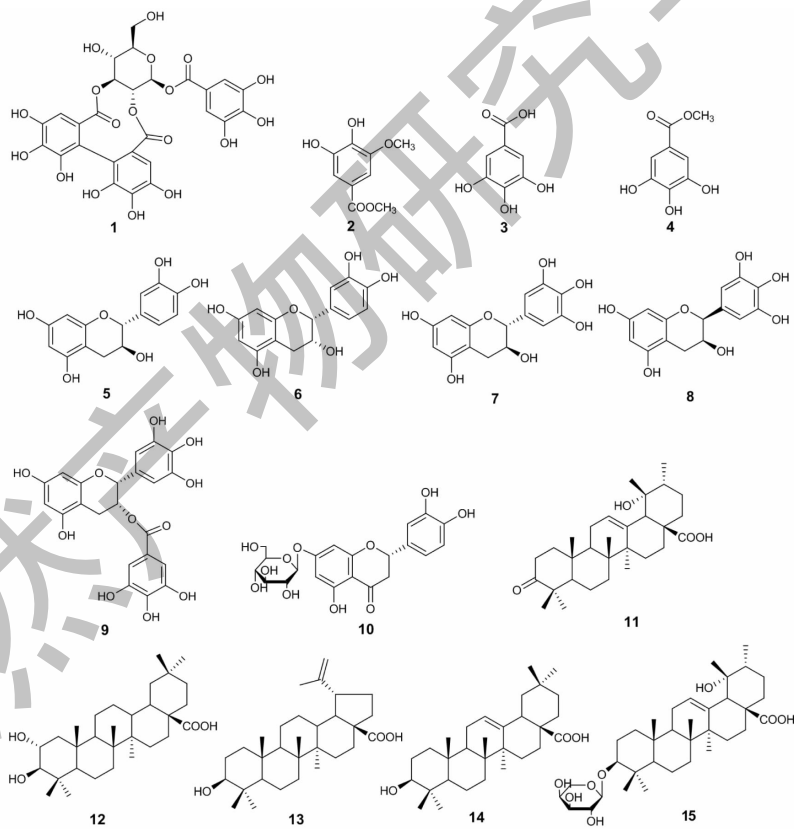
收稿日期: 2019-11-20 接受日期: 2020-05-28

基金项目: 国家自然科学基金 (81573551)

* 通信作者 Tel: 86-451-8669-9347; E-mail: chunjuanyang@126.com

功效,用于血热所致各种出血症,尤适于便血,血痢、崩漏等,也可用于热毒痈肿疮疡、烧伤及湿疹^[1]。地榆因其具有营养价值丰富,药用价值独特、适宜在各地生长等特征,且在我国各地普遍分布,作为一种常用药材被广泛应用^[2]。地榆中主要含有鞣质、皂苷、黄酮等多种化学成分。临床研究表明地榆药材具有抗肿瘤,抑菌消炎,促进烧伤愈合,预防高脂血症,还具有抗凝血活性^[3,4]。地榆素 H-4 是地榆中特有的活性物质,它可以诱导人 HL-60 细胞中人类多聚(ADP-核糖)聚合酶(PARP)裂解相关的 pro-caspase-3 表达减少和 Caspase-3 活性升高,但不影响正常人外周血单核细胞(PBMC)的活性^[5]。没食子酸可特异性抑制脂肪生成,改善胰岛素信号传导,并同时抑制机体的炎症反应和氧化应激反应^[6]。没食子酸甲酯通过 Akt 和 Btk-PLC γ 2-Ca²⁺ 信号传导显着抑制破骨细胞形成。此外,没食子酸甲酯还

可作为顺铂治疗肿瘤的辅助药物,提高癌症治疗的效果^[7,8]。体外和体内研究表明表没食子儿茶素没食子酸酯对乳腺癌 MCF-7 细胞具有抑制作用。此外,没食子儿茶素没食子酸酯可以诱导 G2/M 期细胞的凋亡,在蛋白质水平上抑制 miR-25 表达并提高 PARP, Caspase-3 和 Caspase-9 的表达水平。这表明没食子儿茶素没食子酸酯可以通过下调 miR-25 和与凋亡相关的蛋白的表达来抑制体内肿瘤的生长^[9]。而且,没食子儿茶素没食子酸酯可以抑制 3T3-L1 脂肪细胞的氧化应激和炎症水平,这可能与 NrF2/HO-1 的上调有关^[10]。木犀草素-7-O- β -D-葡萄糖苷不仅能够破坏 STAT3 的核易位,而且还能通过抑制 HEK2 的活性来阻断能量代谢,抑制糖酵解和 Krebs 途径。这表明木犀草素-7-O- β -D-葡萄糖苷可以作为治疗炎症和增生性疾病的潜在候选药物进行开发^[11]。白桦脂酸可以通过调节 AMPK 信号传



	R ₁	R ₂	R ₃
16	galloyl	galloyl	galloyl
17	galloyl	CH ₃	H
18	α -L-arabinofuranosyl	(E)-3,7-dimethyl-7-hydroxy-2-ene-1-oxyl	H
19	α -L-arabinofuranosyl	(3R)-3,7-dimethyloctane-7-hydroxy-1-oxyl	H
20	H	N-butyl	H

图 1 化合物 1~20 的结构式

Fig. 1 The structure of compounds 1-20

导来抑制胰腺细胞的增殖^[12]。1,4,6-tri-*O*-galloyl- β -*D*-glucopyranose 可以抑制 Wnt/ β -catenin 信号通路,并下调 β -catenin 和 Wnt 靶基因(Dkk1、c-Myc、FGF20、NKD1、Survivin)的表达,上调 Caspase-3 和 PARP 的裂解水平,从而抑制直肠癌细胞数目的增长^[13]。在之前的研究中,本课题已分离得到了 10 个三萜类化合物。本研究中,我们继续对地榆的 70% 乙醇提取物的成分进行分离。

1 仪器与材料

硅胶(H,200~300目,青岛海洋化工有限公司,青岛,中国);ODS(COSMOSIL C₁₈-PREP, Nacalaitesque, inc, Japan);硅胶 GF₂₅₄(青岛海洋化工有限公司,青岛,中国);D-101 大孔吸附树脂(天津大钧有限公司,中国)Agilent 1260 series HPLC(美国 Agilent 公司,美国);YMC-Pack ODS-A column(20 mm × 250 mm, 5 μ m)(日本 YMC 股份有限公司,日本);布鲁克 Avance III 核磁共振波谱仪(北京布鲁克科技有限公司,中国);Agilent 6430 QQQ-LC/MS 质谱仪(美国 Agilent 公司,美国);高效液相色谱、质谱所用试剂均为色谱级,其余试剂均为分析级。地榆药材于 2016 年 12 月购买于安国市同义中药饮片有限公司,由黑龙江中医药大学王振月教授鉴定为中药地榆(*S. officinalis*)。样本存放于哈尔滨医科大学药物分析及分析化学教研室(20161202)。

2 提取与分离

干燥地榆根粉碎过 60 目筛,称取 20.0 kg,加入 10 倍量 70% 乙醇溶液,80 °C 加热回流提取 3 次,每次 1h,过滤浓缩,获到乙醇提取物(2 170.3 g)。加适量蒸馏水充分混悬,依次使用等量的乙酸乙酯溶液和正丁醇溶液进行萃取,取上清液,减压浓缩,得到乙酸乙酯萃取物(730.1 g)和正丁醇萃取物(989.7 g)。取正丁醇萃取物过 D101 大孔吸附树脂,用 H₂O、30% EtOH、60% EtOH、90% EtOH 进行系统洗脱,后对其 30% EtOH 提取物通过硅胶柱色谱进行分离,流动相为 CH₂Cl₂-MeOH(100:1→1:1, V/V),共得到 40 个流分(Fr. 1~40)基于薄层色谱分析进行收集。取乙酸乙酯萃取物通过硅胶柱色谱进行分离,流动相为 CH₂Cl₂-MeOH(100:1→1:1, V/V),得到 60 个流分(Fr. 41~110)基于薄层色谱分析收集。Fr. 7 用 ODS 柱进行分离,使用 MeOH-H₂O(10:90→100:0, V/V)梯度洗脱,通过制备型 HPLC-RID 进行纯化,得到化合物 1(31.8 mg)、16(34.2 mg)、化合物 17(105.2 mg)、化合物 18(22.3

mg)和化合物 19(50.7 mg);Fr. 6 用 ODS 柱进行分离,使用 MeOH-H₂O(10:90→100:0, V/V)梯度洗脱,通过制备型 HPLC-RID 进行纯化,得到化合物 2(20.5 mg)和化合物 20(21.3 mg);Fr. 66 用 ODS 柱进行分离,使用 MeOH-H₂O(10:90→100:0, V/V)梯度洗脱,通过制备型 HPLC-RID 进行纯化,得到化合物 3(21.2 mg)和化合物 4(85.1 mg);Fr. 4 用 ODS 柱进行分离,使用 MeOH-H₂O(10:90→100:0, V/V)梯度洗脱,通过制备型 HPLC-RID 进行纯化,得到化合物 5(571.2 mg)、化合物 6(285.1 mg)、化合物 7(62.3 mg)、化合物 8(41.5 mg)和化合物 9(151.0 mg);Fr. 69 用硅胶柱进行分离,使用 CH₂Cl₂-MeOH(100:1→1:1, V/V)梯度洗脱,得到化合物 10(15.2 mg);Fr. 10 用 ODS 柱进行分离,使用 MeOH-H₂O(10:90→100:0, V/V)梯度洗脱,通过制备型 HPLC-RID 进行纯化,得到化合物 11(85.1 mg);Fr. 11 用 ODS 柱进行分离,使用 MeOH-H₂O(10:90→100:0, V/V)梯度洗脱,通过制备型 HPLC-RID 进行纯化,得到化合物 12(85.1 mg);Fr. 58 用 ODS 柱进行分离,使用 MeOH-H₂O(10:90→100:0, V/V)梯度洗脱,通过制备型 HPLC-RID 进行纯化,得到化合物 13(31.8 mg);Fr. 60 用硅胶柱进行分离,使用 CH₂Cl₂-MeOH(100:1→1:1, V/V)梯度洗脱,得到化合物 14(25.2 mg);Fr. 65 用 ODS 柱进行分离,使用 MeOH-H₂O(10:90→100:0, V/V)梯度洗脱,通过制备型 HPLC-RID 进行纯化,得到化合物 15(20.5 mg)。

3 结构鉴定

化合物 1 C₂₇H₂₂O₁₈, 淡黄色粉末;ESI-MS: *m/z* 634 [M-H]⁻; ¹H NMR(400 MHz, CD₃OD) δ : 4.19(1H, m, H-1), 5.36(1H, m, H-2), 5.95(1H, m, H-3), 7.36(1H, d, H-4), 3.60(1H, m, H-6), 3.52(2H, m, H-7), 6.77(6H, s, H-16, 20, 25, 27, 35, 39); ¹³C NMR(100 MHz, CD₃OD) δ : 91.3(C-1), 74.4(C-2), 79.1(C-3), 67.8(C-4), 76.5(C-5), 61.7(C-6), 115.3(C-1'), 126.3(C-2'), 108.0(C-3'), 145.8(C-4'), 137.4(C-5'), 146.0(C-6'), 170.0(C-7'), 115.4(C-1''), 126.9(C-2''), 107.6(C-3''), 145.8(C-4''), 137.5(C-5''), 146.7(C-6''), 171.4(C-7''), 120.3(C-1'''), 110.5(C-2'''), 146.7(C-3'''), 5'''), 140.6(C-4'''), 166.4(C-7''')。以上波谱数据与文献^[14]与地榆素 H-4 的数据基本一致,故鉴定该化合物为地榆素 H-4。

化合物 2 $C_9H_{10}O_5$, 白色无定形粉末, 易溶于甲醇; ESI-MS: m/z 197 $[M-H]^-$; 1H NMR (400 MHz, CD_3OD) δ : 7.19 (1H, d, $J = 1.96$ Hz, H-2), 7.17 (1H, d, $J = 1.96$ Hz, H-6), 3.85 (3H, s, 3-OMe), 3.88 (3H, s, 7-OMe); ^{13}C NMR (100 MHz, CD_3OD) δ : 121.5 (C-1), 106.1 (C-2), 149.2 (C-3), 140.6 (C-4), 146.3 (C-5), 111.9 (C-6), 168.9 (C-7), 52.4 (3-OMe), 56.7 (7-OMe)。以上波谱数据与文献报道^[15] 3,4-二羟基-5-甲氧基苯甲酯的数据基本一致, 故鉴定该化合物为 3,4-二羟基-5-甲氧基苯甲酯。

化合物 3 $C_7H_6O_3$, 白色针状结晶, 易溶于甲醇、乙醇和丙酮; ESI-MS: m/z 171 $[M+H]^+$; 1H NMR (400 MHz, CD_3OD) δ : 7.05 (2H, s, H-2, 6); ^{13}C NMR (100 MHz, CD_3OD) δ : 121.6 (C-1), 109.9 (C-2), 146 (C-3), 139.2 (C-4), 146 (C-5), 109.9 (C-6), 170 (C-7)。以上数据与文献^[16] 报道没食子酸的数据基本一致, 故鉴定该化合物为没食子酸。

化合物 4 $C_8H_8O_3$, 白色针状结晶, 易溶于甲醇、乙醇和丙酮; ESI-MS: m/z 185 $[M+H]^+$; 1H NMR (400 MHz, CD_3OD) δ : 7.04 (2H, s, H-2, 6), 3.79 (3H, s, CH_3); ^{13}C NMR (100 MHz, CD_3OD) δ : 121.5 (C-1), 110.1 (C-2), 146.5 (C-3), 140 (C-4), 146.5 (C-5), 110.1 (C-6), 169.1 (C-7), 52.3 (C-8)。以上数据与文献^[17] 报道没食子酸甲酯的数据基本一致, 故鉴定该化合物为没食子酸甲酯。

化合物 5 $C_{15}H_{14}O_6$, 黄褐色粉末, 易溶于甲醇; ESI-MS: m/z 291 $[M+H]^+$; 1H NMR (400 MHz, CD_3OD) δ : 6.86 (1H, d, $J = 2.0$ Hz, H-2'), 6.78 (1H, d, $J = 7.8$ Hz, H-5'), 6.73 (1H, dd, $J = 2.0, 7.8$ Hz, H-6'), 5.95 (1H, d, $J = 2.1$ Hz, H-6), 5.88 (1H, d, $J = 2.1$ Hz, H-8), 4.58 (1H, d, $J = 7.5$ Hz, H-2), 3.98 (1H, $J = 5.4, 7.5, 13.2$ Hz, H-3), 2.85 (1H, dd, $J = 5.4, 16.4$ Hz, H-4a), 2.53 (1H, dd, $J = 7.8, 16.4$ Hz, H-4b); ^{13}C NMR (100 MHz, CD_3OD) δ : 81.5 (C-2), 67.4 (C-3), 27.1 (C-4), 155.5 (C-5), 94.9 (C-6), 156.2 (C-7), 94.1 (C-8), 156.4 (C-9), 99.4 (C-10), 130.8 (C-1'), 113.9 (C-2'), 144.8 (C-3'), 144.8 (C-4'), 114.7 (C-5'), 118.7 (C-6')。以上波谱数据与文献报道^[18] 儿茶素的数据基本一致, 故鉴定该化合物为儿茶素。

化合物 6 $C_{15}H_{14}O_6$, 黄褐色粉末, 易溶于甲醇; ESI-MS: m/z 291 $[M+H]^+$; 1H NMR (400 MHz,

CD_3OD) δ : 6.83 (1H, d, $J = 1.8$ Hz, H-2'), 6.81 (1H, d, $J = 8.1$ Hz, H-5'), 6.77 (1H, dd, $J = 1.8, 8.1$ Hz, H-6'), 5.96 (1H, d, $J = 2.1$ Hz, H-6), 5.94 (1H, d, $J = 2.1$ Hz, H-8), 4.84 (1H, d, $J = 7.5$ Hz, H-2), 4.20 (1H, sex., $J = 5.4, 7.5, 13.2$ Hz, H-3), 2.86 (1H, dd, $J = 5.4, 16.1$ Hz, H-4a), 2.74 (1H, dd, $J = 8.1, 16.1$ Hz, H-4b); ^{13}C NMR (100 MHz, CD_3OD) δ : 78.5 (C-2), 66.1 (C-3), 27.9 (C-4), 156.0 (C-5), 95.0 (C-6), 156.3 (C-7), 94.5 (C-8), 156.6 (C-9), 98.7 (C-10), 130.9 (C-1'), 113.9 (C-2'), 144.4 (C-3'), 144.5 (C-4'), 114.5 (C-5'), 130.9 (C-6')。以上波谱数据与文献报道^[18] 表儿茶素的数据基本一致, 故鉴定该化合物为表儿茶素。

化合物 7 $C_{15}H_{14}O_7$, 淡黄褐色粉末, 易溶于甲醇; ESI-MS: m/z 307 $[M+H]^+$; 1H NMR (400 MHz, CD_3OD) δ : 6.40 (2H, s, H-2', 6'), 5.92 (1H, d, $J = 2.2$ Hz, H-6), 5.86 (1H, d, $J = 2.2$ Hz, H-8), 4.53 (1H, d, $J = 6.8$ Hz, H-2), 3.97 (1H, sex., $J = 5.2, 6.8, 13.2$ Hz, H-3), 2.82 (1H, dd, $J = 5.2, 16.2$ Hz, H-4a), 2.50 (1H, dd, $J = 6.8, 16.2$ Hz, H-4b); ^{13}C NMR (100 MHz, CD_3OD) δ : 81.5 (C-2), 67.4 (C-3), 26.7 (C-4), 155.4 (C-5), 94.9 (C-6), 156.2 (C-7), 94.1 (C-8), 156.4 (C-9), 99.3 (C-10), 130.2 (C-1'), 105.8 (C-2'), 145.5 (C-3'), 132.6 (C-4'), 145.5 (C-5'), 105.8 (C-6')。以上波谱数据与文献报道^[19] 没食子儿茶素的数据基本一致, 故鉴定该化合物为没食子儿茶素。

化合物 8 $C_{15}H_{14}O_7$, 淡黄褐色粉末, 易溶于甲醇; ESI-MS: m/z 307 $[M+H]^+$; 1H NMR (400 MHz, CD_3OD) δ : 6.40 (2H, s, H-2', 6'), 5.92 (1H, d, $J = 2.2$ Hz, H-6), 5.86 (1H, d, $J = 2.2$ Hz, H-8), 4.53 (1H, d, $J = 6.8$ Hz, H-2), 3.97 (1H, sex., $J = 5.2, 6.8, 13.2$ Hz, H-3), 2.82 (1H, dd, $J = 5.2, 16.2$ Hz, H-4a), 2.50 (1H, dd, $J = 6.8, 16.2$ Hz, H-4b); ^{13}C NMR (100 MHz, CD_3OD) δ : 81.5 (C-2), 67.4 (C-3), 26.7 (C-4), 155.4 (C-5), 94.9 (C-6), 156.2 (C-7), 94.1 (C-8), 156.4 (C-9), 99.4 (C-10), 130.2 (C-1'), 105.8 (C-2'), 145.5 (C-3'), 132.6 (C-4'), 145.5 (C-5'), 105.8 (C-6')。以上波谱数据与文献报道^[20] 表没食子儿茶素的数据基本一致, 故鉴定该化合物为表没食子儿茶素。

化合物 9 $C_{22}H_{18}O_{11}$, 白色粉末, 易溶于甲醇; ESI-MS: m/z 459 $[M+H]^+$; 1H NMR (400 MHz, Pyr-

idine- d_5) δ : 6.71 (2H, s, Gal-H), 6.65 (2H, s, H-2', 6'), 5.28 (2H, d, H-6, 8), 4.65 (1H, br s, H-2), 3.32 (1H, dd, H-4); ^{13}C NMR (100 MHz, Pyridine- d_5) δ : 83.2 (C-2), 68.1 (C-3), 29.0 (C-4), 158.3 (C-5), 96.5 (C-6), 157.2 (C-7), 95.4 (C-8), 158.6 (C-9), 100.8 (C-10), 131.5 (C-1'), 107.5 (C-2'), 147.5 (C-3'), 135.0 (C-4'), 147.5 (C-5'), 107.5 (C-6'), 122.8 (Gal-1), 110.5 (Gal-2), 147.9 (Gal-3), 140.4 (Gal-4), 147.9 (Gal-5), 110.5 (Gal-6), 169.6 (C=O)。以上波谱数据与文献报道^[21]表没食子儿茶素没食子酸酯的数据基本一致,因此鉴别此化合物为表没食子儿茶素没食子酸酯。

化合物 10 $\text{C}_{21}\text{H}_{20}\text{O}_{11}$, 黄色针状结晶;ESI-MS: m/z 449 $[\text{M} + \text{H}]^+$; ^1H NMR (400 MHz, DMSO- d_6) δ : 7.48 (1H, d, $J = 2.4$ Hz, H-2'), 7.45 (1H, dd, $J = 8.2, 2.4$ Hz, H-6'), 6.94 (1H, d, $J = 8.2$ Hz, H-5'), 6.82 (1H, d, $J = 2.0$ Hz, H-8), 6.77 (1H, s, H-3), 6.47 (1H, d, $J = 2.0$ Hz, H-6), 5.12 (1H, d, $J = 7.3$ Hz, H-Glu-1); ^{13}C NMR (100 MHz, DMSO- d_6) δ : 82.8 (C-2), 68.8 (C-3), 28.5 (C-4), 157.6 (C-5), 96.4 (C-6), 157.8 (C-7), 95.6 (C-8), 156.9 (C-9), 100.9 (C-10), 132.2 (C-1'), 116.2 (C-2'), 146.2 (C-3'), 146.2 (C-4'), 115.3 (C-5'), 120.1 (C-6')。以上数据与文献^[22]报道木犀草素-7-*O*- β -D-葡萄糖苷的数据基本一致,故鉴定该化合物为木犀草素-7-*O*- β -D-葡萄糖苷。

化合物 11 $\text{C}_{30}\text{H}_{48}\text{O}_4$, 白色无定形粉末;ESI-MS: m/z 473.4 $[\text{M} + \text{H}]^+$; ^1H NMR (400 MHz, Pyridine- d_5) δ : 5.52 (1H, br. s, H-12), 4.35 (1H, m, H-3), 3.06 (1H, s, H-18), 1.73 (3H, s, H-27), 1.21 (3H, s, H-29), 1.13 (3H, s, H-23), 1.11 (3H, s, H-26), 1.06 (3H, d, $J = 6.0$ Hz, H-30), 1.00 (3H, s, H-24), 0.91 (3H, s, H-25); ^{13}C NMR (100 MHz, Pyridine- d_5) δ : 39 (C-1), 32.6 (C-2), 216.1 (C-3), 47.2 (C-4), 55.3 (C-5), 19.7 (C-6), 34.2 (C-7), 40.2 (C-8), 46.6 (C-9), 36.7 (C-10), 23.8 (C-11), 127.6 (C-12), 139.7 (C-13), 42.5 (C-14), 28.7 (C-15), 26.3 (C-16), 47.2 (C-17), 54.3 (C-18), 72.3 (C-19), 42.4 (C-20), 35.7 (C-21), 74.8 (C-22), 26.7 (C-23), 21.4 (C-24), 14.7 (C-25), 16.7 (C-26), 24.6 (C-27), 179.6 (C-28), 19.1 (C-29), 16.4 (C-30)。以上波谱数据与文献报道^[23]坡模酮酸的数据基本一致,故鉴定该化合物为坡模酮酸。

化合物 12 $\text{C}_{30}\text{H}_{48}\text{O}_3$, 白色无定形粉末,易溶于氯仿、甲醇和吡啶;ESI-MS: m/z 456 $[\text{M}]^+$; ^1H NMR (400 MHz, Pyridine- d_5) δ : 5.45 (1H, t, $J = 3.2$ Hz, H-12), 3.37 (1H, dd, $J = 5.8, 10.3$ Hz, H-3), 3.27 (1H, dd, $J = 4.1, 13.8$ Hz, H-18), 1.26 (3H, s, H-23), 1.23 (3H, s, H-27), 1.06 (3H, s, H-29), 1.00 (3H, s, H-24), 0.97 (3H, s, H-30), 0.94 (3H, s, H-26), 0.93 (3H, s, H-25); ^{13}C NMR (100 MHz, Pyridine- d_5) δ : 47.9 (C-1), 68.3 (C-2), 83.6 (C-3), 39.8 (C-4), 55.7 (C-5), 18.6 (C-6), 33.2 (C-7), 39.7 (C-8), 48.1 (C-9), 38.5 (C-10), 23.6 (C-11), 122.2 (C-12), 144.6 (C-13), 42 (C-14), 28.2 (C-15), 23.5 (C-16), 46.4 (C-17), 42 (C-18), 46.4 (C-19), 30.9 (C-20), 34.2 (C-21), 33.1 (C-22), 29.3 (C-23), 17.5 (C-24), 16.6 (C-25), 17.2 (C-26), 25.9 (C-27), 180 (C-28), 33.2 (C-29), 23.7 (C-30)。以上波谱数据与文献报道^[24]2 α -羟基齐墩果酸的数据基本一致,故鉴定该化合物为2 α -羟基齐墩果酸。

化合物 13 $\text{C}_{30}\text{H}_{48}\text{O}_3$, 白色针状结晶,易溶于甲醇和吡啶;ESI-MS: m/z 457 $[\text{M} + \text{H}]^+$; ^1H NMR (400 MHz, Pyridine- d_5) δ : 4.94 (1H, d, $J = 2.4$ Hz, H-29b), 4.76 (1H, d, $J = 2.4$ Hz, H-29a), 3.45 (1H, t, $J = 8.0$ Hz, H-19), 1.78 (3H, s, H-30), 1.22 (3H, s, H-27), 1.05 (3H, d, $J = 3.6$ Hz, H-23), 1.00 (3H, s, H-25), 0.81 (3H, s, H-24); ^{13}C NMR (100 MHz, Pyridine- d_5) δ : 37.5 (C-1), 26.1 (C-2), 78.1 (C-3), 41.1 (C-4), 55.9 (C-5), 18.8 (C-6), 34.8 (C-7), 42.3 (C-8), 51 (C-9), 37.6 (C-10), 18.8 (C-11), 21.2 (C-12), 38.6 (C-13), 42.9 (C-14), 31.2 (C-15), 32.9 (C-16), 56.6 (C-17), 49.8 (C-18), 47.8 (C-19), 151.3 (C-20), 30.3 (C-21), 39.5 (C-22), 28.7 (C-23), 16.4 (C-24), 16.4 (C-25), 16.4 (C-26), 14.9 (C-27), 178.9 (C-28), 19.5 (C-29), 110 (C-30)。以上数据与文献^[25]报道白桦酯酸的数据基本一致,故鉴定该化合物为白桦酯酸。

化合物 14 $\text{C}_{30}\text{H}_{48}\text{O}_3$, 白色无定形粉末,易溶于氯仿、甲醇和吡啶;ESI-MS: m/z 457 $[\text{M} + \text{H}]^+$; ^1H NMR (400 MHz, Pyridine- d_5) δ : 5.49 (1H, t, $J = 3.2$ Hz, H-12), 3.43 (1H, dd, $J = 5.8, 10.3$ Hz, H-3), 3.30 (1H, dd, $J = 4.1, 13.8$ Hz, H-18), 1.28 (3H, s, H-23), 1.24 (3H, s, H-27), 1.02 (3H, s, H-29), 1.02 (3H, s, H-24), 1.00 (3H, s, H-30), 0.94 (3H, s, H-26), 0.87 (3H, s, H-25); ^{13}C NMR (100 MHz, Pyri-

dine- d_5) δ : 39 (C-1), 28.1 (C-2), 78.1 (C-3), 39.4 (C-4), 55.8 (C-5), 18.8 (C-6), 33.3 (C-7), 39.9 (C-8), 48.2 (C-9), 37.4 (C-10), 23.7 (C-11), 122.6 (C-12), 144.9 (C-13), 42.2 (C-14), 28.4 (C-15), 23.9 (C-16), 46.7 (C-17), 42 (C-18), 46.5 (C-19), 31 (C-20), 34.3 (C-21), 33.3 (C-22), 28.8 (C-23), 16.6 (C-24), 15.6 (C-25), 17.5 (C-26), 26.2 (C-27), 180.3 (C-28), 33.2 (C-29), 23.8 (C-30)。以上数据与文献^[26,27]报道齐墩果酸的数据基本一致,故鉴定该化合物为齐墩果酸。

化合物 15 $C_{35}H_{56}O_8$, 白色无定形粉末, 易溶于甲醇和吡啶; ESI-MS: m/z 627 [M + Na]⁺; ¹H NMR (400 MHz, Pyridine- d_5) δ : 5.61 (1H, brs, H-12), 4.78 (1H, d, $J = 7.0$ Hz, H-1'), 3.34 (1H, dd, $J = 11.7, 4.3$ Hz, H-3), 3.06 (1H, s, H-18), 1.76 (3H, s, H-27), 1.45 (3H, s, H-29), 1.27 (3H, s, H-23), 1.13 (3H, d, $J = 6.7$ Hz, H-30), 1.09 (3H, s, H-26), 0.98 (3H, s, H-24), 0.89 (3H, s, H-25); ¹³C NMR (100 MHz, Pyridine- d_5) δ : 38.9 (C-1), 26.7 (C-2), 88.8 (C-3), 39.6 (C-4), 56 (C-5), 18.7 (C-6), 33.6 (C-7), 40.4 (C-8), 47.8 (C-9), 37 (C-10), 24.1 (C-11), 128.1 (C-12), 140 (C-13), 42.2 (C-14), 29.4 (C-15), 26.4 (C-16), 48.3 (C-17), 54.7 (C-18), 72.5 (C-19), 42.4 (C-20), 27 (C-21), 38.6 (C-22), 28.3 (C-23), 17 (C-24), 15.6 (C-25), 17.2 (C-26), 24.8 (C-27), 180.7 (C-28), 27.2 (C-29), 16.8 (C-30), 107.6 (C-1'), 74.7 (C-2'), 73 (C-3'), 69.6 (C-4'), 66.8 (C-5')。以上数据与文献^[28]报道地榆皂苷 II 的数据基本一致,故鉴定该化合物为地榆皂苷 II。

化合物 16 $C_{28}H_{26}O_{18}$, 黄褐色粉末, 易溶于甲醇; ESI-MS: m/z [M + H]⁺; ¹H NMR (400 MHz, CD₃OD) δ : 4.62 (1H, d, $J = 8$ Hz, H-1), 5.22 (1H, t, $J = 8$ Hz, H-2), 5.94 (1H, t, $J = 8$ Hz, H-3), 3.60-4.00 (2H, m, H-4, 5), 4.50 (1H, br s, H-6), 3.32 (3H, s, -OCH₃), 7.03, 7.06, 7.14 (each 2H, s, Gal-H); ¹³C NMR (100 MHz, CD₃OD) δ : 56.8 (OCH₃), 94.1 (C-1), 74.3 (C-2), 76.5 (C-3), 71.4 (C-4), 76.0 (C-5), 64.3 (C-6), 110.3 (Gal-2, 6), 120.1, 121.1, 121.3 (Gal-1), 139.9 (Gal-4), 146.4, 146.5 (Gal-3, 5), 166.5, 167.6, 168.3 (-COO-)。以上波谱数据与文献报道^[29]1,4,6-三-*O*-没食子酰基- β -*D*-吡喃葡萄糖的数据基本一致,所以鉴定该化合物为 1,

4,6-三-*O*-没食子酰基- β -*D*-吡喃葡萄糖。

化合物 17 $C_{14}H_{18}O_{10}$, 淡黄色粉末, 易溶于甲醇; ESI-MS: m/z 347 [M + H]⁺; ¹H NMR (400 MHz, Pyridine- d_5) δ : 7.19 (2H, s, H-15, 19), 4.96 (1H, dd, $J = 1.6, 12.0$ Hz, H-11a), 4.65 (1H, dd, $J = 5.6, 12.0$ Hz, H-11b), 4.18 (1H, d, $J = 7.6$ Hz, H-6), 3.98 (3H, s, H-21); ¹³C NMR (100 MHz, Pyridine- d_5) δ : 105.5 (C-1), 74.7 (C-2), 75.2 (C-3), 71.2 (C-4), 78.1 (C-5), 64.5 (C-6), 56.4 (C-7), 167.1 (C-8), 121.0 (C-9), 110.0 (C-10, 14), 149.4 (C-11, 13), 135.2 (C-12)。以上波谱数据与文献报道^[30]methyl-6-*O*-galloyl- β -*D*-glucopyranoside 的数据基本一致,因此鉴定此化合物为 methyl-6-*O*-galloyl- β -*D*-glucopyranoside。

化合物 18 $C_{21}H_{38}O_{11}$, 白色粉末, 易溶于甲醇; ¹H NMR (400 MHz, CD₃OD) δ : 5.40 (1H, t, $J = 6.5$, H-2), 4.35 (1H, dd, $J = 11.9, 6.3$, H-1), 4.32 (1H, d, $J = 6.8$, H-1''), 4.29 (1H, d, $J = 7.8$, H-1'), 4.21 (1H, dd, $J = 11.9, 7.8$, H-1), 4.09 (1H, dd, $J = 11.2, 1.9$, H-6'), 3.73 (1H, dd, $J = 11.2, 5.4$, H-6'), 2.05 (1H, t, $J = 7.1$, H-4); ¹³C NMR (100 MHz, CD₃OD) δ : 63.1 (C-1), 121.5 (C-2), 142.2 (C-3), 41.1 (C-4), 23.4 (C-5), 44.3 (C-6), 71.4 (C-7), 29.2 (C-8), 29.2 (C-9), 16.5 (C-10), 102.8 (C-1'), 75.1 (C-2'), 78.1 (C-3'), 71.4 (C-4'), 76.8 (C-5'), 68.1 (C-6'), 102.8 (C-1''), 72.0 (C-2''), 79.0 (C-3''), 68.1 (C-4''), 66.4 (C-5'')。以上波谱数据与文献报道^[31](*E*)-7-hydroxy-3,7-dimethyl-2-octenyl-6-*O*- α -*L*-arabinofuranosyl- β -*D*-glucopyranoside 的数据基本一致,故鉴定该化合物为 (*E*)-7-hydroxy-3,7-dimethyl-2-octenyl-6-*O*- α -*L*-arabinofuranosyl- β -*D*-glucopyranoside。

化合物 19 $C_{21}H_{40}O_{11}$, 白色粉末, 易溶于甲醇; ¹H NMR (400 MHz, CD₃OD) δ : 4.96 (1H, d, $J = 1.26$, H-1''), 4.25 (1H, d, $J = 7.9$, H-1'), 3.55 (1H, dt, $J = 9.4, 7.1$, H-1), 4.02 (1H, dd, $J = 11.2, 2.4$, H-6'), 3.61 (1H, dd, $J = 11.2, 6.0$, H-6'); ¹³C NMR (100 MHz, CD₃OD) δ : 69.4 (C-1), 37.9 (C-2), 30.9 (C-3), 38.9 (C-4), 22.7 (C-5), 45.1 (C-6), 71.5 (C-7), 29.2 (C-8), 29.3 (C-9), 20.1 (C-10), 104.5 (C-1'), 75.1 (C-2'), 78.1 (C-3'), 72.1 (C-4'), 76.7 (C-5'), 68.2 (C-6'), 110.0 (C-1''), 83.2 (C-2''), 79.0 (C-3''), 86.0 (C-4''), 63.1 (C-

5'')。以上波谱数据与文献报道^[32] 7-hydroxy-3,7-dimethyloctyl-6-O- α -L-arabinofuranosyl- β -D-glucopyranoside 的数据基本一致,故鉴定该化合物为 7-hydroxy-3,7-dimethyloctyl-6-O- α -L-arabinofuranosyl- β -D-glucopyranoside。

化合物 20 C₁₀H₂₀O₆,白色无定形粉末,硫酸显色为黑色;ESI-MS: m/z 235 [M + Na]⁺; ¹H NMR (400 MHz, CD₃OD) δ : 4.75 (1H, d, J = 3.8, H-1), 3.67 (1H, dt, J = 9.9, 6.4, H-1'), 3.42 (2H, dt, J = 9.9, 6.4, H-2', H-3'), 1.57 (1H, m, H-4'); ¹³C NMR (100 MHz, CD₃OD) δ : 100.1 (C-1), 73.6 (C-2), 75.1 (C-3), 71.9 (C-4), 73.6 (C-5), 62.7 (C-6), 68.8 (C-1'), 32.8 (C-2'), 20.5 (C-3'), 14.3 (C-4')。以上波谱数据与文献报道^[33] n-Butyl- β -D-glucopyranoside 的数据基本一致,故鉴定该化合物为 n-Butyl- β -D-glucopyranoside。

4 结论

本研究通过 70% 的乙醇对地榆进行提取后,对乙酸乙酯和正丁醇萃取层进行分离,结合 D101 大孔吸附树脂,硅胶柱色谱,ODS 柱色谱,制备型高效液相等手段进行分离纯化,以核磁共振和质谱确定化合物的结构。从地榆的 70% 乙醇提取物中共分离得到 20 个单体化合物,其中 4 个酚酸类化合物、6 个黄酮类化合物、5 个皂苷类化合物和 5 个糖苷类化合物。其中化合物 9、20 为首次从地榆属植物中分得,该研究对地榆中主要化学成分进行了进一步确证。

参考文献

- Hu J, Song Y, Li H, et al. Cytotoxic triterpene glycosides from the roots of *Sanguisorba officinalis* [J]. Arch Pharm Res, 2014, 38:984-990.
- Xia HM, Sun LL, Sun JY, et al. Progress on chemical ingredient and pharmacological activity of *Sanguisorba officinalis* L. [J]. Food Drug(食物与药品), 2009, 11(7):67-69.
- Sun W, Zhang ZL, Liu X, et al. Terpene glycosides from the roots of *Sanguisorba officinalis* L. and their hemostatic activities [J]. Molecules, 2012, 17:7629-7636.
- Zhang S, Liu X, Zhang Z, et al. Isolation and identification of the phenolic compounds from the roots of *Sanguisorba officinalis* L. and their antioxidant activities [J]. Molecules, 2012, 17:13917-13922.
- Chen LG, Huang WT, Lee LT, et al. Ellagitannins from *Terminalia catappa* L. induced apoptosis in HL-60 cells [J]. Toxicol In Vitro, 2009, 23:603-609.
- Dludla PV, Nkambule BB, Jack B, et al. Inflammation and oxidative stress in an obese state and the protective effects of gallic acid [J]. Nutrients, 2018, 11(1):23.
- Kim H, Lee G, Sohn SH, et al. Immunotherapy with methyl gallate, an inhibitor of Treg cell migration, enhances the anti-cancer effect of cisplatin therapy [J]. Korean J Physiol Pharmacol, 2016, 20(3):261-268.
- Baek JM, Kim JY, Lee CH, et al. Methyl gallate inhibits osteoclast formation and function by suppressing akt and btk-PLC γ 2-Ca²⁺ signaling and prevents lipopolysaccharide-induced bone loss [J]. Int J Mol Sci, 2017, 18(3):581.
- Zan L, Chen Q, Zhang L, et al. Epigallocatechin gallate (EGCG) suppresses growth and tumorigenicity in breast cancer cells by downregulation of miR-25 [J]. Bioengineered, 2019, 10(1):374-382.
- Kudelaite M, Tang WJ, He WN, et al. Effect of epigallocatechin-3-gallate on oxidative stress and inflammation in 3T3-L1 adipocytes [J]. J Shanghai Jiaotong Univ: Med Sci(上海交通大学学报:医学版), 2018, 38:1289-1294.
- Palombo R, Caporali S, Falconi M, et al. Luteolin-7-O- β -D-Glucoside inhibits cellular energy production interacting with HEK2 in keratinocytes [J]. Int J Mol Sci, 2019, 20:2689.
- Sun L, Cao J, Chen K, et al. Betulinic acid inhibits stemness and EMT of pancreatic cancer cells via activation of AMPK signaling [J]. Int J Oncol, 2019 Jan, 54(1):98-110.
- Li W, Yang CJ, Wang LQ, et al. A tannin compound from *Sanguisorba officinalis* blocks Wnt/ β -catenin signaling pathway and induces apoptosis of colorectal cancer cells [J]. Chin Med(中华医学杂志), 2019, 14:22-35.
- Kapustina II, Makar'eva TN, Kalinovskii AI, et al. Glycosides and thymidine from the mollusk *Cryptochiton stelleri* [J]. Chem Nat Compd +, 2005, 41(1):109-110.
- Su XD, Ali I, Arooj M, et al. Chemical constituents from *Sanguisorba officinalis* L. and their inhibitory effects on LPS-stimulated pro-inflammatory cytokine production in bone marrow-derived dendritic cells [J]. Arch Pharm Res, 2018, 41:497-505.
- Ma X, Liu Y, Shi Y. Phenolic derivatives with free-radical-scavenging activities from *Ixeridium gracile* (DC.) Shih [J]. Chem Biodivers, 2007, 4:2172-2181.
- Wang ZY, Wang LN, Qiu L, et al. Isolation and identification of phenolic constituents of *Sanguisorbae Radix* [J]. Chin J Exp Tradit Med Form(中国实验方剂学杂志), 2017, 23(8):82-85.
- Wang GK, Zhang J, Yu Y, et al. Chemical constituents of ethyl acetate fraction from *Dioscorea bulbifera* [J]. Chin Pharm J

- (中国药学杂志), 2018, 53: 1815-1820.
- 19 Cheliger. Study on chemical constituents medicine *Tetraena mongolica* Maxim [D]. Tongliao: National University of the Inner Mongol (内蒙古民族大学), 2017.
 - 20 Ji XZ, Wang QH, Wu J. Chemical constituents from roots of *Eucommia ulmoides* Oliv [J]. Bio Chem Eng (生物化工), 2017, 3(3): 40-42.
 - 21 Zhou YL, Dong BS, Zhang FQ, et al. Study on chemical constituents of cooked puerh [J]. Yunnan Chem Tech (云南化工), 2009, 36(2): 10-13.
 - 22 Ma X, Liu Y, Shi Y. Phenolic derivatives with free-radical-scavenging activities from *Ixeridium gracile* (DC.) Shih [J]. Chem Biodivers, 2007, 4: 2172-2181.
 - 23 Liang Q, Gong Z, Min Z. Studies on triterpenes from *Elephantopus scaber* [J]. Chem Pharm Bull, 2007, 42: 494-496.
 - 24 Yang L, Lin J, Zhou B, et al. Activity of compounds from *Taxillus sutchuenensis* as inhibitors of HCV NS3 serine protease [J]. Nat Prod Res, 2017, 31: 487-491.
 - 25 Irungu B, Orwa J, Gruhonjic A, et al. Constituents of the roots and leaves of *Ekebergia capensis* and their potential antiplasmodial and cytotoxic activities [J]. Molecules, 2014, 19: 14235-14246.
 - 26 Qin W, Wu X, Zhao J, et al. Triterpenoid glycosides from leaves of *Ilex cornuta* Lindl [J]. Phytochemistry, 1986, 25: 913-916.
 - 27 Nomizu K, Hashida K, Makino R, et al. Antioxidants from steamed used tea leaves and their reaction behavior [J]. Biosci Biotechnol Biochem, 2008, 72: 1682-1689.
 - 28 Zdero C, Bohlmann F, King R, et al. Guaianolides and labdanes from *Brickellia* species [J]. Phytochemistry, 1991, 30: 1591-1595.
 - 29 Lee SH, Tanaka T, Nonaka G, et al. Alloose gallates from *Euphorbia fischeriana* [J]. Phytochemistry, 1991, 30: 1251-1253.
 - 30 Tanaka T, Nonaka G, Nishioka I. Tannins and related compounds. XVI. isolation and characterization of six methyl glucoside gallates and a gallic acid glucoside gallate from *Sanguisorba officinalis* L. [J]. Chem Pharm Bull, 1984, 2(1): 117-121.
 - 31 Su XD, Ali I, Arooj M, et al. Chemical constituents from *Sanguisorba officinalis* L. and their inhibitory effects on LPS-stimulated pro-inflammatory cytokine production in bone marrow-derived dendritic cells [J]. Arch Pharm Res, 2018, 41: 497-505.
 - 32 Su XD, Guo RH, Li HX, et al. Anti-allergic inflammatory components from *Sanguisorba officinalis* L. [J]. Bioorg Med Chem Lett, 2018, 28: 2210-2216.
 - 33 Song YN, Shibuya M, Ebizuka Y, et al. Identification of plant factors inducing virulence gene expression in *Agrobacterium tumefaciens* [J]. Chem Pharm Bull, 1991, 39: 2347-2350.
-
- (上接第 131 页)
- 20 Wang JN, Ming J, Tian Y, et al. Extraction and antioxidant activities of flavonoids from *Coreopsis tinctoria* Nutt tea [J]. Food Mach (食品与机械), 2018, 34(9): 162-166.
 - 21 Nie C, Zhao ZY, Liu LY, et al. Effect of extraction parameters on the yield of flavanones and polymethoxy flavones from citrus orange peel and antioxidant activity of the extract [J]. Food Ferment Ind (食品与发酵工业), 2018, 44(1): 158-165.
 - 22 Jiang H, Wang MP, Tian T, et al. An optimized method for infusing *Chrysanthemum* tea [J]. Food Sci (食品科学), 2011, 32(22): 152-155.
 - 23 Yu XY, Fang JX, Xiang Q, et al. Extraction optimization by response surface methodology and HPLC analysis of organic acids from pixian broad bean paste [J]. Food Sci (食品科学), 2019, 40(4): 286-291.
 - 24 Xu HB, Li B, Tang ZS, et al. Optimization of extraction of total flavonoids from *hovenia acerba* and its xanthine oxidase inhibitory activity [J]. Nat Prod Res Dev (天然产物研究与开发), 2019, 31: 595-600.
 - 25 Ci ML, Chen Y, Zhang X, et al. Optimization of extraction process of blueberry anthocyanin by response surface methodology and its antioxidant activity [J]. Food Ind (食品工业), 2014, 35(4): 39-44.
 - 26 Gil MI, Tomás-Barberán FA, Hess-Pierce B, et al. Antioxidant activity of pomegranate juice and its relationship with phenolic composition and processing [J]. J Agr Food Chem, 2000, 48: 4581-4589.
 - 27 Dong PX, Ya L, Xiao M, et al. Natural antioxidants in foods and medicinal plants: extraction, assessment and resources [J]. Int J Mol Sci, 2017, 18(1): 96-101.
 - 28 Tan LH, Yang ZF, Zhang D, et al. Research progress of antioxidant components and evaluation methods of traditional Chinese medicine [J]. Asia Pac Tradit Med (亚太传统医药), 2017, 13(10): 35-37.