

UPLC-Q-TOF-MS/MS 分析凉粉草多酚组分的化学成分

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摘要:岭南特色药材凉粉草(*Mesona chinensis* Benth.)具有很好的消暑、清热、解毒等功能,被称为“仙草”。本研究采用超高效液相色谱串联四极杆飞行时间质谱(UPLC-Q-TOF-MS/MS),选用Waters Acquity UPLC BEH C₁₈色谱柱(2.1 mm × 100 mm, 1.7 μm);流动相为0.1%甲酸乙腈溶液(A)-0.1%甲酸水溶液(B),梯度洗脱;电喷雾离子源(ESI),负离子监测。共鉴定出51个化合物,包括40个酚类、3个萜类和8个其它化合物,其中1个苯丙酸、1个苯丙酸二聚体、1个苯丙酸三聚体、3个黄酮和3个萜类首次在凉粉草植物中被发现。

关键词:凉粉草;多酚;UPLC-Q-TOF-MS/MS

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Analysis of polyphenols from *Mesona chinensis* by UPLC-Q-TOF-MS/MS

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Abstract: *Mesona chinensis* Benth., a characteristic herb of south of China, is known as “Immortal herb” because of its excellent functions of heat elimination, heat clearing, detoxification and so on. Ultra-performance liquid chromatography coupled with quadrupole time-of-flight mass/mass spectrometry (UPLC-Q-TOF-MS) was selected for the study. A BEH C₁₈ column (2.1 mm × 100 mm, 1.7 μm) was used with the mobile phase of eluent (A) MeCN containing 0.1% formic acid and eluent B 0.1% formic acid in water. Q-TOF-MS and electrospray ion (ESI) source were applied for the analysis under the negative ions. Fifty-one compounds were identified including 40 polyphenols, 3 terpenoids, and 8 other compounds, whereas 1 phenylpropionic acids, 1 lignan dimers, 2 lignan trimmers, 3 flavonoids and 3 terpenoids were firstly identified in *Mesona chinensis* Benth.

Key words: *Mesona chinensis* Benth.; polyphenols; UPLC-Q-TOF-MS/MS

凉粉草(*Mesona chinensis* Benth.),是唇形科凉粉草属草本植物,分布在印度东北部,东南亚和中国台湾、福建、广东、广西和云南。凉粉草作为一种中药材,被誉为“仙草”,具有收敛、甘甜、清寒、清热、凉血、解毒的功效^[1],并且已经被开发为食品、天然保健品甚至是临床药品。现有研究证明,它的根、茎、叶及花各个组织含有大量的生物活性物质,包括多糖、黄酮、萜类、维生素和酚类^[2],具有很高的营养和医药保健价值。

酚类化合物对植物的生长起着重要的作用,对人体的各个器官也有十分强的保护作用。据报道,有学者通过研究发现凉粉草含有降血糖和高血压物质,包括β-谷甾醇、豆甾醇、α-和β-淀粉蛋白、齐墩果酸、马兜铃酸和β-谷甾醇糖苷^[3]。同时,也有学者从凉粉草的水提液中鉴定出山奈酚、芹菜素、咖啡酸、原儿茶酸、丁香酸、香草酸、对羟基苯甲酸、齐墩果酸和熊果酸九种活性化合物^[4],其中咖啡酸、原儿茶酸、山奈酚等被认为是凉粉草抗氧化活性的重要组成部分,包括降低LDL氧化和炎症,从而提供预防心血管疾病的保护。

超高效液相色谱串联四极杆飞行时间质谱(UPLC-Q-TOF-MS/MS)技术,具有灵敏度高、分辨率

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高、分离效率高、分析时间短等优点^[5],同时还能得到化合物的分子量、元素组成以及离子碎片等信息,可快速确定化合物的结构,在药物分析领域得到越来越广泛的应用。目前尚不完全了解凉粉草多酚的成分,本实验采用 UPLC-Q-TOF-MS/MS 技术对凉粉草中的化学成分进行快速、全面地定性分析,为凉粉草酚类化学成分的研究提供重要参考。

1 材料、试剂与仪器

1.1 材料

凉粉草样品于 2019 年 6 月采自福建省龙岩市,经暨南大学张英副教授鉴定为凉粉草 (*Mesona chinensis* Benth.)。

1.2 主要试剂

乙腈(色谱纯)购自美国 Sigma 公司、甲酸(色谱纯)购自美国 Sigma 公司、甲醇(色谱纯)瑞典 Oceanpak 公司;单体对照品:caffeic acid、quercetin-3-O-rutinoside、isoquercetin、quercetin 和 kaempferol 国药集团化学试剂有限公司(纯度≥98.0%);AB-8 大孔树脂购自东鸿化工有限公司;纯水为 Milli-Q 系统纯化水。其他试剂均为分析纯。

1.3 仪器

超高效液相色谱-四级杆-飞行时间串联质谱联用仪 (Nexera UHPLC LC-30A + AB SCIEX Triple TOF™ 5600+);PHS-3B 雷磁 pH 计(上海精密科学仪器有限公司);ML-104 电子天平(上海梅特勒-托利多仪器有限公司);Milli-Q Advantage A10 纯水系统;普通层析柱(40 mm×60 cm)。

2 方法

2.1 凉粉草多酚提取

将凉粉草全草进行清洗后,在阴凉通风处风干,

切碎,干燥保存。准确称重凉粉草 60 g,用 1.5 L 去离子水在微沸状态下提取 3 次,每次 2 h^[6]。参照 Li 等^[7]和 Bai 等^[8]纯化方法,略有改动,采用 AB-8 大孔树脂进行纯化,按质量比 1/40(样品/填料)进行上样,先用去离子水洗至洗脱液无多糖和蛋白质等杂质,再用 75% 乙醇进行多酚化合物的洗脱,将洗脱液富集、浓缩和冷冻干燥得多酚提取物。

2.2 供试品和对照品溶液的制备

将凉粉草多酚提取物和对照品用甲醇充分溶解,稀释至 50 μg/mL,过 0.22 μm 有机微孔滤膜即得,供 UPLC-Q-TOF-MS/MS 分析。

2.3 UPLC-Q-TOF-MS/MS 分析条件

2.3.1 色谱条件

色谱柱为 Waters Acquity UPLC BEH C₁₈ 色谱柱(2.1 mm×100 mm,1.7 μm),流速为 0.3 mL/min。柱温为 40 °C,进样量为 3 μL。洗脱液为 A,0.1% 的甲酸乙腈溶液和 B,0.1% 的甲酸水溶液。梯度洗脱条件:0~1 min,5%→10% A;1~5 min,10%→30% A;5~8 min,30% A;8~9 min,30%→50% A;9~12 min,50%→60% A;12~20 min,60%→80% A;20~21 min,80%→90% A;21~22 min,90%→95% A^[9]。

2.3.2 质谱条件

以负 ESI 模式运行,并配备 DuoSpray™ 离子源 (AB Sciex, 加利福尼亚州福斯特城)。MS-MS 检测器的条件如下:离子喷雾电压为 4.5 kV;离子源加热器,40 °C;帘气,35 psi;雾化气体(GS1),55 psi;TIS 气体(GS2),55 psi;碰撞能量(CE),35 eV;碰撞能量扩散(CES),15 eV。质量范围设置为 *m/z* 100~2 000^[9]。采用 Peak View 分析软件和 Scifinder 网

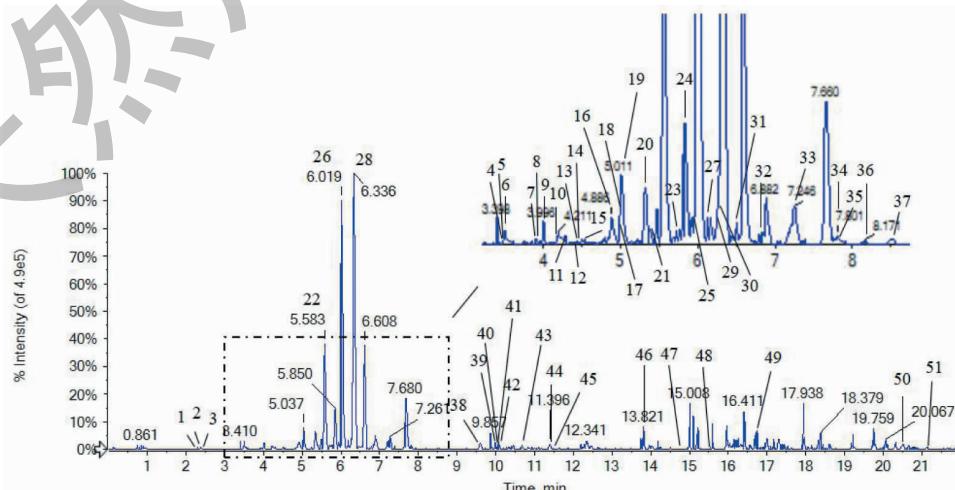


图 1 凉粉草多酚负离子模式下的总离子流图

Fig. 1 The total ion chromatograms of polyphenols from *M. chinensis* in negative ion modes

络数据库进行检索和分析。

3 结果与讨论

本研究采用 UPLC-Q-TOF-MS/MS 技术测定凉粉草总酚提取物成分的保留时间、前体离子、MS/MS 产物离子。凉粉草多酚样品在负离子模式下的

总离子电流色谱图如图 1 所示,共鉴定出 51 个化合物,包括 8 个苯丙酸、3 个苯丙酸二聚体、4 个苯丙酸三聚体、1 个苯丙酸四聚体、17 个黄酮类、7 个酚酸类、3 个萜类和 8 个其它化合物。相关化合物物质谱信息和结构见表 1 和图 2。

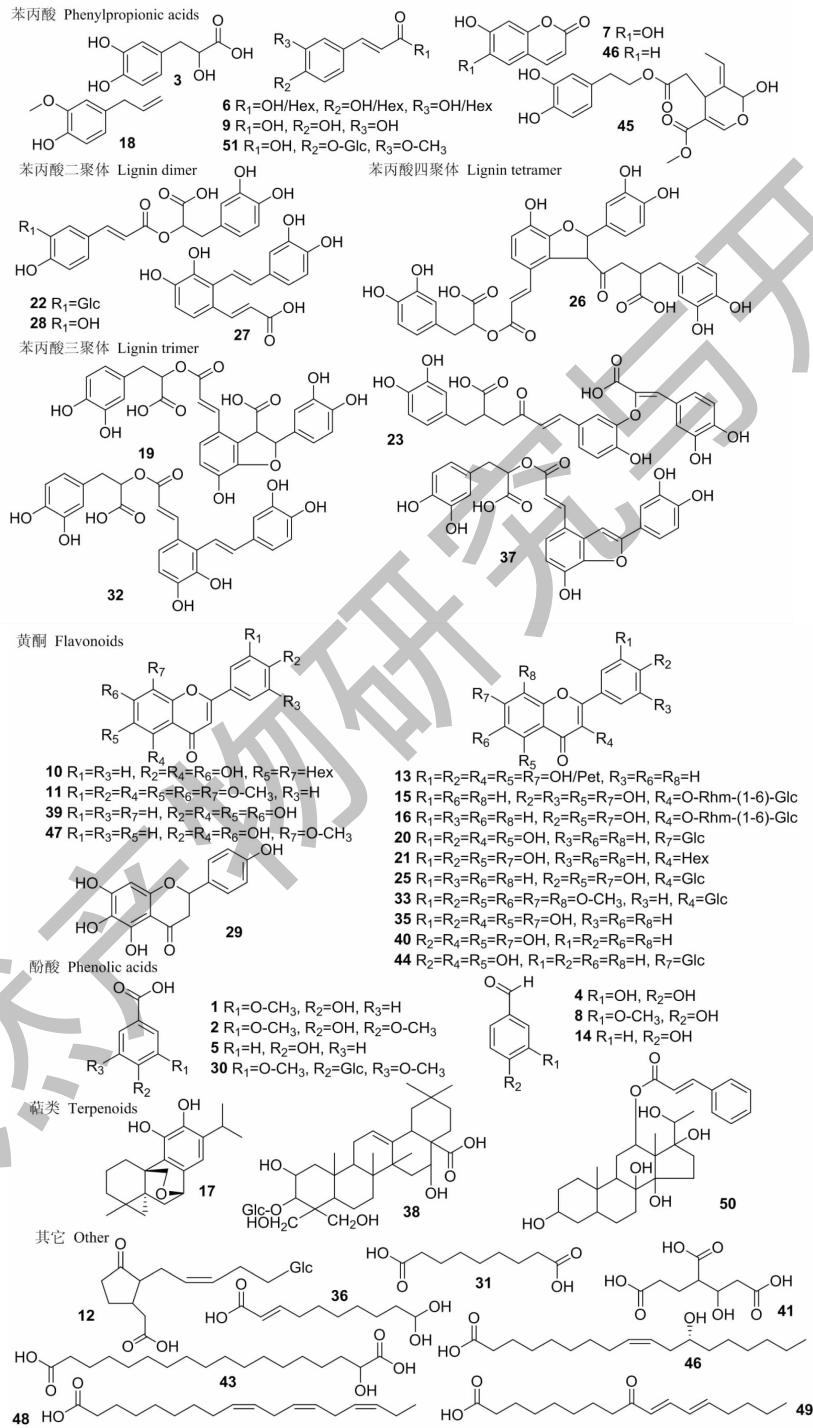


图 2 凉粉草多酚化合物的结构

Fig. 2 Structures of polyphenols from *M. chinensis*

表 1 凉粉草 UPLC-Q-TOF-MS/MS 分析结果
Table 1 UPLC-Q-TOF-MS/MS analysis results of *Meisona chinensis*

序号 No.	类别 Classify	出峰时间 t_R (min)	[M-H] ⁻ 观测值 Observed [M-H] ⁻ (m/z)	分子式 Formula	碎片离子 Fragment ions (MS/MS)	化合物 Compound	参考文献 Ref.
3	苯丙酸	2.39	197.045 8	C ₉ H ₁₀ O ₅	197.807 9, 179.037 4, 151.040 5, 135.045 9, 123.046 8, 107.911 2	丹参素 Danshensu	10
6		3.48	341.087 1	C ₁₅ H ₁₈ O ₉	341.087 2, 281.069 7, 251.057 8, 221.053 3, 179.034 5, 135.045 2, 134.036 4	咖啡酰己糖苷 Caffeoyl hexoside ^{FI}	11
7		3.90	177.019 4	C ₉ H ₆ O ₄	177.019 4, 160.843 2, 149.023 5, 133.031 1, 121.029 8, 109.029 5, 105.035 6	6,7-二羟基香豆素 6,7-Dihydroxycoumarin	11
9		3.99	179.035 5	C ₉ H ₈ O ₄	179.034 3, 161.033 3, 136.047 8, 135.044 9, 134.036 8, 117.034 1, 107.050 1	咖啡酸 Caffeic acid	对照品
18		4.93	163.040 5	C ₁₀ H ₁₂ O ₂	162.839 0, 145.889 6, 119.050 3, 117.034 7, 104.028 7	丁香酚 Eugenol	12
42		10.07	161.024 5	C ₉ H ₆ O ₃	161.024 9, 143.899 4, 133.029 3, 132.040 3, 124.890 9, 116.930 0, 105.012 4	7-羟基香豆素 Umbelliferone	13
45		11.49	377.172 6	C ₁₉ H ₂₂ O ₈	377.097 3, 341.197 2, 284.883 2, 174.956 7	Oleoeuropein aglycone	14
51		21.21	369.242 0	C ₁₆ H ₁₈ O ₁₀	337.215 6, 311.167 2, 193.122 0, 163.112 0, 161.096 3	阿魏酸葡萄糖苷 Ferulic acid-4-O-glucuronide	15
22	苯丙酸 二聚体	5.53	521.128 9	C ₂₄ H ₂₆ O ₁₃	521.130 5, 359.077 0, 197.045 5, 161.025 1, 135.045 2	异迷迭香酸苷 Rosmarinic acid-3-O-glucoside	15
27		6.16	313.071 4	C ₁₇ H ₁₄ O ₆	314.094 7, 239.948 4, 174.955 9, 151.040 2, 121.035 8	丹酚酸 F Salvianolic acid F ^{FI}	16
28		6.27	359.076 3	C ₁₈ H ₁₆ O ₈	359.203 8, 359.076 0, 197.044 5, 161.023 9, 135.044 6	迷迭香酸 Rosmarinic acid	15
19	苯丙酸 三聚体	5.00	537.102 6	C ₂₇ H ₂₂ O ₁₂	537.104 1, 357.060 8, 339.049 7, 313.071 3, 295.060 1, 229.012 2	紫草酸 Lithospermic acid	17
23		5.71	537.102 2	C ₂₇ H ₂₂ O ₁₂	469.833 2, 339.049 6, 295.060 0, 185.022 0	丹酚酸 H Salvianolic acid H ^{FI}	18
32		6.75	493.111 4	C ₂₆ H ₂₂ O ₁₀	493.111 9, 447.097 5, 313.074 7, 295.058 9, 135.241 5	丹酚酸 A Salvianolic acid A	19
37		8.52	491.096 4	C ₂₆ H ₂₀ O ₁₀	491.174 2, 311.054 3, 293.044 0, 265.049 5, 179.032 7, 135.045 6	丹酚酸 C Salvianolic acid C	11
26	苯丙酸 四聚体	5.92	717.145 3	C ₃₆ H ₃₀ O ₁₆	673.159 1, 537.104 2, 519.093 0, 493.112 3, 383.071 8, 339.049 5, 321.039 4, 295.059 7, 179.032 0	丹酚酸 B Salvianolic acid B	17
10	黄酮	4.18	593.150 2	C ₂₇ H ₃₀ O ₁₅	503.116 5, 473.107 8, 353.065 2, 297.078 8	Apigenin-6,8-di-C-hexoside ^{FI}	20
11		4.24	401.239 1	C ₂₁ H ₂₂ O ₈	365.000 0, 343.000 0, 295.000 0, 269.000 0, 203.000 0, 109.000 0	川陈皮素 Nobiletin ^{FI}	21
13		4.37	433.207 1	C ₂₀ H ₁₈ O ₁₁	387.197 5, 295.905 2, 235.920 9, 189.127 0	槲皮素戊聚糖 Quercetin-pentoside	13
15		4.51	609.146 0	C ₂₇ H ₃₀ O ₁₆	463.090 8, 446.085 9, 299.016 9	芦丁 Quercetin-3-O-rutinoside	对照品
16		4.88	593.149 5	C ₂₇ H ₃₀ O ₁₅	593.208 4, 447.093 6, 327.047 2, 285.039 5, 284.031 6	Aromadendrin-3-O-rutinoside	12
20		5.32	463.086 9	C ₂₁ H ₂₀ O ₁₂	463.086 4, 301.033 7, 300.026 1, 271.023 6, 255.028 7, 151.002 3	槲皮黄昔 Quercimeritin	22
21		5.34	463.087 5	C ₂₁ H ₂₀ O ₁₂	463.087 6, 301.034 6, 300.026 6, 271.023 7, 255.028 1	Quercetin-3-O-hexoside	23
24		5.83	447.092 0	C ₂₁ H ₂₀ O ₁₁	285.038 3, 284.031 2, 255.028 2, 227.033 7, 151.002 3	Isomer of kaempferol-3-O-glucoside	24

续表1(Continued Tab. 1)

序号 No.	类别 Classify	出峰时间 <i>t_R</i> (min)	[M-H] ⁻ 观测值 Observed [M-H] ⁻ (<i>m/z</i>)	分子式 Formula	碎片离子 (MS/MS)	化合物 Compound	参考文献 Ref.
25		5.85	447.092 2	C ₂₁ H ₂₀ O ₁₁	285.039 9, 284.031 9, 255.029 5, 227.034 0	紫云英苷 Astragalin	24
29		6.27	287.149 7	C ₁₅ H ₁₂ O ₆	287.147 1, 269.141 1, 259.061 0, 227.129 1, 209.117 7, 125.021 1	红花素 Carthamidin	25
33		7.25	579.315 6	C ₂₇ H ₃₁ O ₁₄	533.312 1, 453.302 1, 399.254 0, 337.259 0	Nobiletin-3-O-β-D-glucoside ^{FI}	26
34		7.81	301.034 9	C ₁₅ H ₁₀ O ₇	273.039 6, 255.031 7, 178.997 1, 151.002 9, 121.029 7	异槲皮素 Isoquercetin	对照品
35		7.83	301.034 9	C ₁₅ H ₁₀ O ₇	301.976 3, 255.227 5, 178.997 4, 151.005 1, 179.027 9, 121.028 7	槲皮素 Quercetin	对照品
39		9.97	285.040 0	C ₁₅ H ₁₀ O ₆	285.039 7, 257.045 6, 239.033 8, 229.049 4, 211.040 5	野黄芩素 Scutellarein	27
40		9.99	285.040 2	C ₁₅ H ₁₀ O ₆	285.039 4, 239.03 34, 229.049 8, 211.038 9	山奈酚 Kaempferol	对照品
44		11.39	447.132 0	C ₂₁ H ₂₀ O ₁₁	447.133 3, 431.102 9, 242.942 0, 174.954 8, 149.001 1	Kaempferol-7-O-glucoside	28
47		14.72	299.185 6	C ₁₆ H ₁₂ O ₆	284.031 0, 231.949 3, 185.010 2, 181.159 3, 142.943 0	Isoscutellarein-8-methyl ether	27
1	酚酸	2.32	167.035 4	C ₈ H ₈ O ₄	167.035 1, 149.025 9, 137.025 7, 123.045 6, 121.030 1, 109.030 5, 108.022 5	香草酸 Vanillic acid	29
2		2.39	197.045 8	C ₉ H ₁₀ O ₅	197.807 6, 153.900 7, 135.047 1, 123.046 2	丁香酸 Syringic acid	30
4		3.41	137.025 1	C ₇ H ₆ O ₃	137.455 4, 137.026 4, 136.018 0, 124.019 7, 119.016 6, 109.029 8, 108.023 4	原儿茶醛 Protocatechuic aldehyde	22
5		3.42	138.031 7	C ₇ H ₆ O ₃	137.026 0, 119.014 8, 109.031 7, 108.022 7	对苯酚甲酸 4-Hydroxybenzoic acid	31
8		3.98	151.040 5	C ₈ H ₈ O ₃	151.039 6, 121.004 2, 109.030 0, 108.021 9, 107.062 1	香草醛 Vanillin	21
14		4.37	121.030 1	C ₇ H ₆ O ₂	121.028 7, 108.021 9	对羟基苯甲醛 <i>p</i> -Hydroxybenzaldehyde	21
30		6.29	359.076 4	C ₁₅ H ₂₀ O ₁₀	359.076 9, 197.045 1, 179.096 3, 161.024 2, 137.032 5, 133.029 9	丁香酸葡萄糖苷 Syringic acid glucoside	15
17	萜类	4.89	315.050 3	C ₂₀ H ₂₈ O ₃	315.055 5, 261.838 6, 254.055 0, 181.065 9, 153.071 0, 127.057 3	20-去氧鼠尾草酚 20-Deoxocarnosol ^{FI}	16
38		9.59	695.400 0	C ₃₇ H ₆₀ O ₁₂	649.395 8, 487.341 6, 384.885 3, 346.905 4	3-O- <i>D</i> -glucopyranosyl platycodigenin methyl ester ^{FI}	32
50		20.44	513.308 9	C ₃₀ H ₄₂ O ₇	511.665 8, 445.112 2, 377.865 9, 319.823 9, 235.969 4, 164.930 4	Marstanacigenin A ^{FI}	32
12	其它	4.28	387.164 9	C ₁₈ H ₂₈ O ₉	387.164 9, 341.107 2, 207.102 9, 163.109 7, 113.024 2	Tuberonic acid glucoside	23
31		6.40	187.097 8	C ₉ H ₁₆ O ₄	187.100 1, 169.088 7, 142.942 2, 125.098 2, 124.705 0, 123.081 2	壬二酸 Azelaic acid	26
36		8.16	201.113 3	C ₁₀ H ₁₈ O ₄	201.113 0, 183.100 7, 164.834 1, 147.893 3, 140.116 2, 139.112 7, 137.097 4	(E)-10,10-Dihydroxy-2-decanoic acid	33
41		10.07	205.050 9	C ₇ H ₁₀ O ₇	205.049 6, 177.054 3, 161.023 6, 135.044 1, 125.872 5, 105.033 3	Homoisocitric acid	34
43		10.66	329.232 5	C ₁₈ H ₃₄ O ₅	330.238 8, 293.210 4, 229.144 9, 211.132 8, 171.102 2, 139.110 5	Hydroxy octadecanedioic acid	35
46		13.86	297.152 5	C ₁₈ H ₃₄ O ₃	239.074 9, 225.057 7, 184.015 0, 183.011 5, 170.006 1, 119.050 6	蓖麻油酸 Ricinoleic acid	32

续表1(Continued Tab. 1)

序号 No.	类别 Classify	出峰时间 t_R (min)	[M-H] ⁻ 观测值 Observed [M-H] ⁻ (m/z)	分子式 Formula	碎片离子 (MS/MS)	化合物 Compound	参考文献 Ref.
48		15.56	277.1438	$C_{18}H_{30}O_2$	277.1438, 233.1535, 208.9391, 181.8960, 147.0073, 134.0380, 127.1118	α -亚麻酸 α -Linolenic acid	36
49		16.78	293.1786	$C_{18}H_{30}O_3$	293.1784, 221.1579, 182.2146	(E, E)-9-Oxoctadeca-10, 12-dienoic acid	26

注:FI:首次在凉粉草植物中发现。Note:FI:Firstly identified in *M. chinensis*.

3.1 苯丙酸

在凉粉草多酚样品中,化合物3、6、7、9、18、42、45和51属于苯丙酸类化合物,分别是丹参素^[10]、咖啡酰己糖苷^[11]、6,7-二羟基香豆素^[11]、咖啡酸、丁香酚^[12]、7-羟基香豆素^[13]、oleoeuropein aglycone^[14]和阿魏酸葡萄糖苷^[15]。以咖啡酸和咖啡酰己糖苷为例,二者质谱裂解途径见图3。咖啡酰己糖苷母离子[M-H]⁻为 m/z 341.0871,再失去一个己糖得到碎片 m/z 179.0343,这是咖啡酸的母离子。咖啡酸进一步裂解,丢失一个羟基得到 m/z 161.0333, m/z 135.0449则是裂解一个羧基后得到的离子碎

片^[11]。

3.2 苯丙酸二聚体

以异迷迭香酸苷^[15]和迷迭香酸^[15]为苯丙酸二聚体的裂解例子,裂解情况见图4,异迷迭香酸苷在二级离子碎片中存在 m/z 179.0343,表现为异迷迭香酸苷脱去一个己糖,得到迷迭香酸(m/z 359.0770)。迷迭香酸进一步裂解,得到离子峰为 m/z 197.0455的丹参素碎片和 m/z 161.0239的碎片,再丢失一个羰基得到离子碎片 m/z 135.0446^[15]。同时,化合物27初步鉴定为丹酚酸F^[16]。

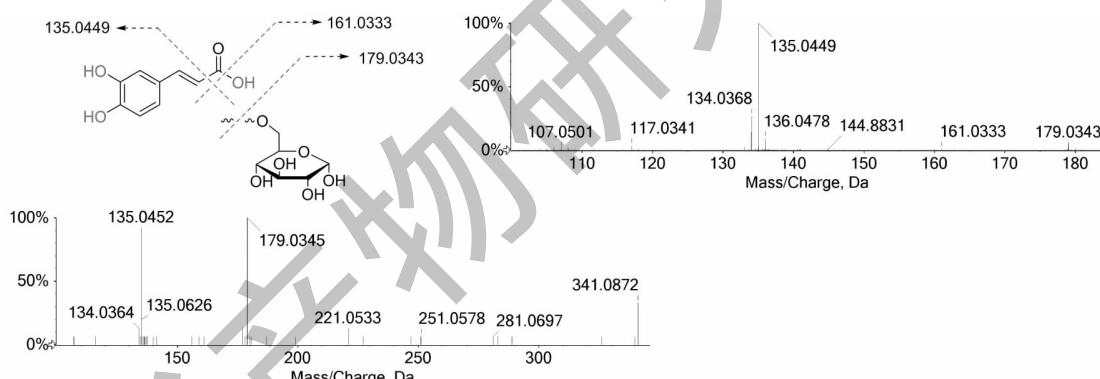


图3 咖啡酸和咖啡酰己糖苷的质谱裂解途径图
Fig. 3 Fragmentation pathways of caffeic acid and caffeoyl hexoside

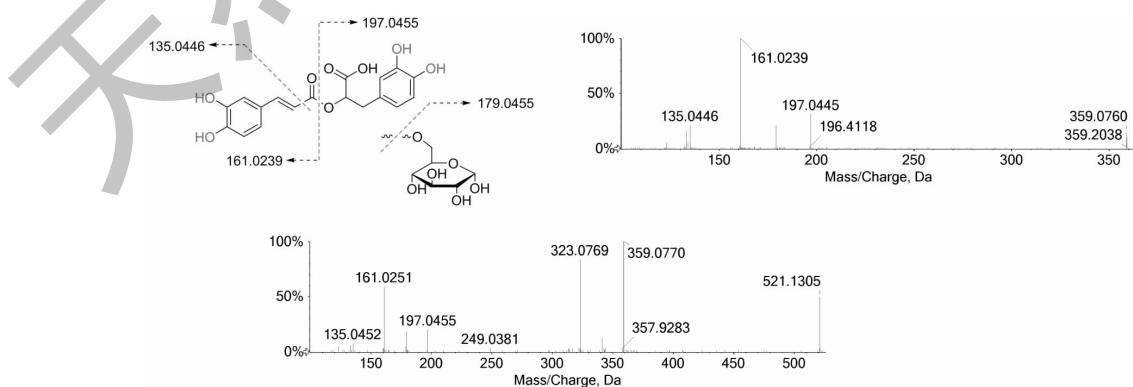


图4 迷迭香酸己糖苷和迷迭香酸的质谱裂解途径图

Fig. 4 Fragmentation pathways of rosmarinic acid-*O*-hexoside and rosmarinic acid

3.3 苯丙酸三聚体

4个三聚体在凉粉草多酚样品中被鉴定,分别是紫草酸^[17]、丹酚酸H^[18]、丹酚酸A^[19]和丹酚酸C^[11]。丹酚酸C的裂解规律如图5,[M-H]⁻为m/z 491.1742,类似迷迭香酸,在丹参素分子处发生裂

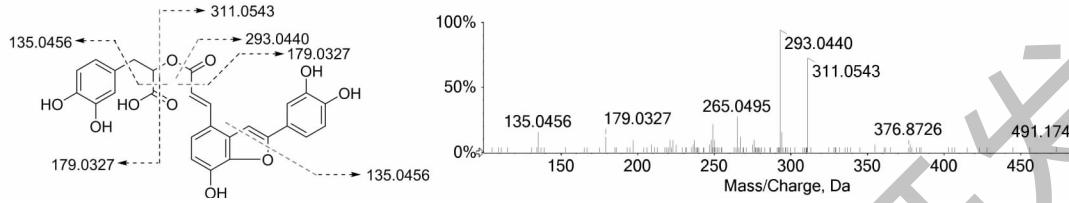


图5 丹酚酸C的质谱裂解途径图

Fig. 5 Fragmentation pathways of salvianolic acid C

3.4 苯丙酸四聚体

化合物26的母离子[M-H]⁻为m/z 717.1453,分子式C₃₆H₃₀O₁₆,MS/MS离子产物有m/z 519.093

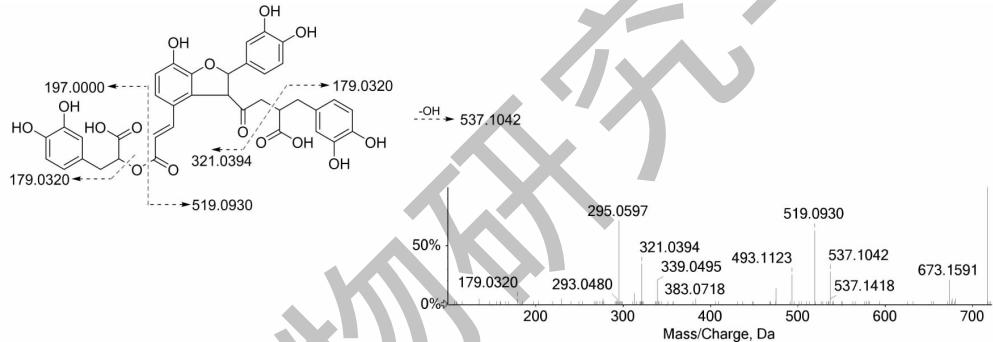


图6 丹酚酸B的质谱裂解途径图

Fig. 6 Fragmentation pathways of salvianolic acid C

3.5 黄酮

一共有4个黄酮类化合物被鉴定。化合物10通过二级离子碎片m/z 503.1165、473.1078和297.0788,初步鉴定为apigenin-6,8-di-C-hexoside^[20]。化合物11、39和47分别被鉴定为nobiletin、scutellarein和isoscutellarein-8-methyl ether。

12个黄酮醇类化合物在凉粉草多酚中被鉴定。化合物13、15、20、21、34和35都有槲皮素的二级特征离子m/z 301,部分还有m/z 295或者255,是槲皮素和它的衍生物,分别被鉴定为槲皮素戊聚糖^[13]、芦丁、槲皮黄苷^[22]、quercetin-3-O-hexoside^[23]、异槲皮素和槲皮素。化合物16、24、25、40和44中,几乎都有m/z 285、255和239的特征离子,初步鉴定为aromadendrin-3-O-rutinoside^[12]、isomer of kaempferol-3-O-glucoside^[24]、紫云英苷^[24]、山奈酚和kaempferol-7-O-glucoside^[28]。化合物33的母离子[M-H]⁻为

解得到m/z 179.0327和311.0543,进一步裂解羧基得到m/z 135.0456和179.0327。同时,因碰撞碎裂产生的子离子还有m/z 293.0440和265.0495^[11]。

0、339.0495、321.0394和179.0320等,初步鉴定为丹酚酸B^[17],质谱裂解途径见图6。

m/z 579.3156,母离子丢失一个葡萄糖得到m/z 399.2540,初步鉴定为nobiletin-3-O-β-D-glucoside。

1个二氢黄酮类被鉴定。化合物29的母离子[M-H]⁻为m/z 287.1497,二级碎片m/z 269.1411、259.0610、227.1291和125.0211,初步鉴定为红花素^[25]。

3.6 酚酸类

一共有7个酚酸类化合物被鉴定。化合物1丢失一个羧基,生成一个特征离子m/z 123.0456,同时拥有特征离子m/z 149.0259、137.0257、121.0301和108.0225,初步鉴定为香草酸^[29]。化合物2的母离子[M-H]⁻为m/z 197.0458,分子式为C₉H₁₀O₅,母离子丢失一个羧基得到m/z 153.9007,再失去一个甲氧基得到m/z 123.0462,初步鉴定为丁香酸^[30]。化合物4和5都拥有的特征离子m/z 119,是丢失一个羟基得到的离子,分别被鉴定为原儿茶

醛^[22]和对苯酚甲酸^[31]。化合物 8 分子式为 C₈H₈O₃,失去一个甲氧基得到离子 m/z 121.004 2,初步鉴定为香草醛^[21]。化合物 14 被鉴定为对羟基苯甲醛^[21]。母离子 [M-H]⁻ 为 m/z 359.076 4 的化合物 30,失去一个葡萄糖得到 m/z 197.045 1,再失去一个羧基得到 m/z 137.032 5,初步鉴定为丁香酸葡萄糖苷^[15]。

3.7 薯类

化合物 17、38 和 50 分别被鉴定为 20-去氧鼠尾草酚^[16]、3-O-D-glucopyranosyl platycodigenin methyl ester^[32] 和 marstenacigenin A^[32], 相关液质数据和结构式见表 1 和图 2。

3.8 其它

化合物 12、31、36、41、43、46、48 和 49 分别被鉴定为 tuberonic acid glucoside^[23]、壬二酸^[26]、(E)-10, 10-dihydroxy-2-decanoic acid^[33]、homoisocitric acid^[34]、hydroxy octadecanedioic acid^[35]、蓖麻油酸^[32]、 α -亚麻酸^[36] 和 (E, E)-9-oxooctadeca-10, 12-dienoic acid^[26], 相关液质数据和结构式见表 1 和图 2。

4 结论

本研究建立了 UPLC-Q-TOF-MS/MS 技术对凉粉草多酚组分进行分析的方法,通过分析质谱原始数据,包括:相对分子质量、保留时间、元素组成、碎片离子等信息,结合对照品、数据库和文献报道分析鉴定,鉴定并推测出 51 个化合物,证实了凉粉草多酚组分主要为黄酮类和苯丙酸及其多聚体组成,其中咖啡酰己糖苷、丹酚酸 F、丹酚酸 H、apigenin-6,8-di-C-hexoside、川陈皮素、nobiletin-3-O- β -D-glucoside、20-去氧鼠尾草酚、3-O-D-glucopyranosyl platycodigenin methyl ester、marstenacigenin A 尚未在凉粉草相关文献中被报道,本研究认为该 9 个化合物在凉粉草中首次被发现。研究结果为凉粉草药效物质基础研究及开发利用奠定基础,对进一步开发新药具有重要的指导意义。

参考文献

- Flora of China Editorial Committee of Chinese Academy of Sciences. Flora of China (中国植物志) [M]. Beijing: Science Press, 1995, 66:547.
- Shyu MH, Kao TC, Yen GC. Hsian-tsao (*Mesona procumbens* Hemsl.) prevents against rat liver fibrosis induced by CCl₄ via inhibition of hepatic stellate cells activation [J]. Food Chem Toxicol, 2008, 46:3707-3713.
- Yen GC, Hung CY. Effects of alkaline and heat treatment on antioxidative activity and total phenolics of extracts from Hsian-tsao (*Mesona procumbens* Hemsl.) [J]. Food Res Int, 2000, 33:487-492.
- Huang HC, Chuang SH, Wu YC, et al. Hypolipidaemic function of Hsian-tsao tea (*Mesona procumbens* Hemsl.): working mechanisms and active components [J]. J Funct Food, 2016, 26:217-227.
- Yuan ZY, Luo LW, Chen NH, et al. Rapid analysis of chemical constituents of Lily Bulbil by UPLC-Q-TOF-MS and the antitumor activity of diosgenin [J]. Nat Prod Res Dev (天然产物研究与开发), 2019, 31:808-813.
- Huang LY, Huang HY, Wang YY, et al. Total phenolic, flavonoid contents and antioxidant capacities of aqueous extract of 16 common edible flowers [J]. Sci Technol Food Ind (食品工业科技), 2017, 38(4):353-356.
- Li YC, Li BX, Lv YF, et al. Study on purification process of polyphenols from blueberry leaves with AB-8 macroporous resins [J]. Sci Technol Food Ind (食品工业科技), 2012, 33(20):258-261.
- Bai WM, Huang GS, Kong WB, et al. Adsorption separation of olive polyphenols from olive oil processing effluent by AB-8 macroporous resin [J]. China Oils Fats (中国油脂), 2015, 40(1):74-77.
- Zheng JX, Tian WY, Yang C, et al. Identification of flavonoids in *Plumula nelumbinis* and evaluation of their antioxidant properties from different habitats [J]. Ind Crop Prod, 2019, 127:36-45.
- Xin S, Jie Z, Xu W, et al. Rapid screening and quantitative determination of active components in Qing-Hua-Yu-Re-Formula using UHPLC-Q-TOF/MS and HPLC-UV [J]. J Anal Methods Chem, 2018, 2018:8535127.
- Contreras MDM, Algieri F, Rodriguez-Nogales A, et al. Phytochemical profiling of anti-inflammatory *Lavandula* extracts via RP-HPLC-DAD-QTOF-MS and -MS/MS: Assessment of their qualitative and quantitative differences [J]. Electrophoresis, 2017, 39:9-10.
- Pandey R, Chandra P, Srivastava M, et al. Simultaneous quantitative determination of multiple bioactive markers in *Ocimum sanctum* obtained from different locations and its marketed herbal formulations using UPLC-ESI-MS/MS combined with principal component analysis [J]. Phytochem Anal, 2015, 26:383-394.
- Zhuang B, Bi ZM, Wang YZ, et al. Chemical profiling and quantitation of bioactive compounds in *Platycladi Cacumen* by UPLC-Q-TOF-MS/MS and UPLC-DAD [J]. J Pharm Biomed Anal, 2018, 154:207.
- Dai J, Mumper RJ. Plant phenolics: extraction, analysis and their antioxidant and anticancer properties [J]. Molecules, 2010, 15:7313-7352.

- 15 Ren Q, Wang YL, Wang ML, et al. Screening and identification of the metabolites in rat urine and feces after oral administration of *Lycopus lucidus* Turcz extract by UHPLC-Q-TOF-MS mass spectrometry [J]. *J Chromatogr B*, 2016, 1027:64-73.
- 16 Yang ST, Wu X, Rui W, et al. UPLC/Q-TOF-MS analysis for identification of hydrophilic phenolics and lipophilic diterpenoids from *Radix Salviae Miltiorrhizae* [J]. *Acta Chromatogr*, 2015, 1(4):1-18.
- 17 Chen Y, Zhang N, Ma J, et al. A Platelet/CMC coupled with offline UPLC-QTOF-MS/MS for screening antiplatelet activity components from aqueous extract of Danshen [J]. *J Pharm Biomed Anal*, 2016, 117:178-183.
- 18 Chen XF, Lou ZY, Zhang H, et al. Identification of multiple components in Guanxinning injection using hydrophilic interaction liquid chromatography/time-of-flight mass spectrometry and reversed-phase liquid chromatography/time-of-flight mass spectrometry [J]. *Rapid Commun Mass Spectrom*, 2011, 25:1661-1674.
- 19 Cai HD, Su SL, Li YH, et al. Danshen can interact with intestinal bacteria from normal and chronic renal failure rats [J]. *Biomed Pharmacother*, 2019, 109:1758-1771.
- 20 Tsamo AT, Ndibewu PP, Dakora FD. Phytochemical profile of seeds from 21 Bambara groundnut landraces via UPLC-qTOF-MS [J]. *Food Res Int*, 2018, 112:160-168.
- 21 Lin LZ, Yang QY, Zhao K, et al. Identification of the free phenolic profile of Adlay bran by UPLC-QTOF-MS/MS and inhibitory mechanisms of phenolic acids against xanthine oxidase [J]. *Food Chem*, 2018, 253:108-118.
- 22 Wei FH, Chen MT, Luo CH, et al. Developing an Absorption-Based quality control method for Hu-Gan-Kang-Yuan capsules by UFLC-QTOF-MS/MS screening and HPLC-DAD quantitative determination [J]. *Molecules*, 2016, 21 (5): 592.
- 23 Zhang L, Xu L, Ye YH, et al. Phytochemical profiles and screening of α -glucosidase inhibitors of four *Acer* species leaves with ultra-filtration combined with UPLC-QTOF-MS/MS [J]. *Ind Crop Prod*, 2019, 129:156-168.
- 24 Xu JY, Yu YL, Shi RY, et al. Organ-specific metabolic shifts of flavonoids in *Scutellaria baicalensis* at different growth and development stages [J]. *Molecules*, 2018, 23 (2):428.
- 25 Hong CC, Chang C, Zhang H, et al. Identification and characterization of polyphenols in different varieties of *Camellia oleifera* seed cakes by UPLC-QTOF-MS [J]. *Food Res Int*, 2019, 126:108614.
- 26 Wang YR, Wang CZ, Li HQ, et al. Discovery of the potential biomarkers for discrimination between *Hedyotis diffusa* and *Hedyotis corymbosa* by UPLC-QTOF/MS metabolome analysis [J]. *Molecules*, 2018, 23 (7):1525.
- 27 Wong TL, An YQ, Yan BC, et al. Comprehensive quantitative analysis of Chinese patent drug YinHuang Drop Pill by ultra high-performance liquid chromatography quadrupole time of flight mass spectrometry [J]. *J Pharm Biomed Anal*, 2016, 125:415-426.
- 28 Karar EGM, Kuhnert N. UPLC-ESI-Q-TOF-MS/MS characterization of phenolics from *Crataegus monogyna* and *Crataegus laevigata* (Hawthorn) leaves, fruits and their herbal derived drops (*Crataegutt Tropfen*) [J]. *J Chem Biol Ther*, 2016, 1:102.
- 29 Kumar B, Pandey R. HPLC-QTOF-MS/MS-based rapid screening of phenolics and triterpenic acids in leaf extracts of *Ocimum* species and their interspecies variation [J]. *J Liq Chromatogr Relat Technol*, 2016, 39:225-238.
- 30 Bai S, Li P, Liu J, et al. A UFLC-MS/MS method for the simultaneous determination of eight bioactive constituents from red wine and dealcoholized red wine in rat plasma: Application to a comparative pharmacokinetic study [J]. *Biomed Chromatogr*, 2018, 33 (3):e4437.
- 31 Cocuron JC, Casas MI, Yang F, et al. Beyond the wall: High-throughput quantification of plant soluble and cell-wall bound phenolics by liquid chromatography tandem mass spectrometry [J]. *J Chromatogr A*, 2019, 1589:93-104.
- 32 Wang CZ, Zhang NQ, Wang ZZ, et al. Nontargeted metabolomic analysis of four different parts of *Platycodon grandiflorum* grown in northeast China [J]. *Molecules*, 2017, 22 (8): 1280.
- 33 Leyva-Jimenez FJ, Lozano-Sanchez S, Borras-Linares I, et al. Potential antimicrobial activity of honey phenolic compounds against Gram positive and Gram negative bacteria [J]. *LWT*, 2019, 101:236-245.
- 34 Fu YY, Shan MQ, Hu MH, et al. Chemical profiling of Banxia-Baizhu-Tianma decoction by ultra-fast liquid chromatography with tandem mass spectrometry [J]. *J Pharm Biomed Anal*, 2019, 174:595-607.
- 35 Farag MA, Weigend M, Luebert F, et al. Phytochemical, phylogenetic, and anti-inflammatory evaluation of 43 *Urtica* accessions (stinging nettle) based on UPLC-Q-TOF-MS metabolomic profiles [J]. *Phytochemistry*, 2013, 96: 170-183.
- 36 Lee YH, Kim B, Kim S, et al. Characterization of metabolite profiles from the leaves of green perilla (*Perilla frutescens*) by ultra high performance liquid chromatography coupled with electrospray ionization quadrupole time-of-flight mass spectrometry and screening for their antioxidant properties [J]. *J Food Drug Anal*, 2017, 25:776-788.